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D-2-Hydroxyglutarate producing neo-enzymatic activity inversely correlates with frequency of the type of isocitrate dehydrogenase 1 mutations found in glioma

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Abstract

Background: IDH mutations frequently occur in diffuse gliomas and result in a neo-enzymatic activity that results in reduction of α -ketoglutarate to D-2-hydroxyglutarate. In gliomas, the frequency of IDH1 mutations in codon 132 increases in the order R132L-R132S-R132G-R132C-R132H with R132H constituting more than 90% of all IDH1 mutations.

Results: We determined the levels of D-2-hydroxyglutarate in glioma tissues with IDH1 mutations. D-2-hydroxyglutarate levels increased in the order of R132H-R132C-R132S/R132G/R132L. We expressed and purified IDH1 wild type and mutant protein for biochemical characterization. Enzyme kinetics of mutant IDH protein correlated well with D-2-hydroxyglutarate production in cells with R132H exhibiting the highest and R132L the lowest K_M for α -ketoglutarate. Addition of D-2-hydroxyglutarate to the medium of cell lines revealed an inhibitory effect at higher concentrations. Migration of LN229 increased at lower D-2-hydroxyglutarate concentrations while higher concentrations showed no effect.

Conclusion: These findings may suggest natural selection against the rare IDH1R132 mutations in human glioma due to toxicity caused by high levels of D-2-hydroxyglutarate.

Keywords: IDH1, 2-HG, D-2-hydroxyglutarate, Enzymatic activity, K_M , Astrocytoma, Oligodendroglioma

Background

Mutations in isocitrate dehydrogenases 1 or 2 (IDH1, IDH2) occur approximately in 75% of diffuse astrocytomas and oligodendroglial tumors [1-3], in 20% of acute myeloid leukemia (AML) [4,5], 50% of chondrosarcoma [6,7], 20% of intrahepatic cholangiocarcinoma [8] and in 20% of angioimmunoblastic T-cell lymphoma [9]. Besides overall frequency, also the type of IDH mutations differs in these tumor entities. In astrocytoma and oligodendroglioma more than 90% of all IDH mutations are of the IDH1R132H type [2], with the second most frequent type (approximately 4% of mutations) being

IDH1R132C. The remaining mutations split into other very rare IDH1 and IDH2 mutations. Quite differently, IDH2R140Q mutations are most frequent in AML [5,10]. In chondrosarcoma and intrahepatic cholangiocarcinoma, the R132C alteration in IDH1 is the most frequent mutation [6,8], while in angioimmunoblastic T-cell lymphoma mutations in IDH2 are predominant [9]. Common to all IDH mutations is a neo-enzymatic activity of the mutated proteins that results in reduction of α -ketoglutarate (α -KG) to D-2-hydroxyglutarate (2-HG) with consumption of NADPH. The neo-enzymatic activity of mutated IDH proteins is believed to constitute the major tumorigenic mechanism of this alteration due to the inhibitory effect of 2-HG on α -KG dependent dioxygenases. Among these, the TET hydroxylases are affected resulting in a severely altered methylation pattern of DNA [11,12] conferring an unknown advantage to these tumor cells.

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In astrocytoma and oligodendroglioma, we detected higher 2-HG levels in rare IDH1 mutations than in the most common IDH1R132H alteration [13]. A similar observation was made for IDH2, with cells carrying mutations affecting R172 exhibiting higher 2-HG concentrations than those with mutations in R140 [14]. These findings suggest different enzyme activity grades for the variant IDH mutants.

To explore the potential role of different mutation types in the formation of astrocytomas and oligodendroglial tumors, we determined the enzyme kinetics of wild type IDH1 and with the mutations R132H, R132C, R132G, R132S, R132L and R100Q followed by quantification of 2-HG in transfected cells and in human tumor tissues, and examined the effects on viability.

Methods

FFPE samples, DNA extraction and IDH1 sequencing

The collection of IDH1 mutant tumors consist out of previously published specimen [1,2,15] and tumors collected at the neuropathology department of the pathological institute of Heidelberg.

The DNA extractions from FFPE samples were performed by hand using Invisorb Genomic DNA Kit II (Invitex, Berlin, Germany) or semi-automated using Maxwell® 16 FFPE Plus LEV DNA Purification Kit (Promega, Madison, USA) following the manufacturer's protocol.

The sequencing of published specimen was performed as described in the earlier publications. Our most recent probes were sequenced by generating a fragment of 212 bp length spanning the catalytic domain of IDH1 including codon 100 and 132 using 60 ng each of the sense primer IDH1-JM_f TGATGAGAAGAGGGTTGAGGA and the antisense primer IDH1-JM_r GCAAATCACAT TATTGCCAAC. PCR using standard buffer conditions, 20 ng of DNA and GoTaq DNA Polymerase (Promega, Madison, USA) employed 35 cycles with denaturing at 95°C for 30 s, annealing at 57°C for 40 s and extension at 72°C for 50 s in a total volume of 15 µl.

A total of 2 µl of the PCR amplification product was submitted to the sequencing reaction using the BigDye Terminator v3.1 Sequencing Kit (Applied Biosystems, Foster City, USA). Twenty-five cycles were performed employing 12 ng of the sense primer IDH1-JM_f TGATGAGAAGAGGGTTGAGGA, with denaturing at 95°C for 30 s, annealing at 57°C for 15 s and extension at 60°C for 240 s. A second round of sequencing analysis was performed using the antisense primer IDH1-JM_r GCAAATCACATTATTGCCAAC and the sequencing reaction conditions as described above. Sequences were determined using the semi-automated sequencer (ABI 3100 Genetic Analyzer, Applied Biosystems, Foster City) and the Sequence Pilot version 3.1 (JSI-Medisys, Kippenheim, Germany) software.

IDH1 mutant generation and cloning

IDH1 mutants were generated using the site directed mutagenesis method. Therefore, primers for each mutation were created and used on IDH1 wt cDNA in pDONR221 (DKFZ clone repository). Each mutation was confirmed by Sanger sequencing using the same procedure as described for the FFPE samples. The pDONR221 clones were used for all further LR-reactions into the described destination vectors. LR-reactions were performed following the manufacturer's protocol (Invitrogen, Carlsbad, USA).

IDH1 protein purification

To purify the different IDH1 proteins, the cDNAs were transferred into pDEST15 (Invitrogen, Carlsbad, USA), an *E. coli* expression vector containing a N-terminal GST tag (Invitrogen, Carlsbad, USA). The pDEST15 vectors were then transfected into *E. coli* expression strain KRX (Promega, Madison, USA). The *E. coli* were then streaked out on lysogeny broth (LB)-plates containing Ampicillin (100 µg/ml, Sigma-Aldrich, St. Luis, USA) and incubated at 37°C overnight. Six clones from each construct were transferred into 6 ml liquid LB containing 100 µg/ml Ampicillin and grown over night at 37°C in an orbital shaker (220 rpm). From these overnight cultures we used aliquots to prepare two small cultures (10 ml) for induction experiments. We only induced one culture from each colony following the manufacturer's protocol (KRX protocol, Promega, Madison, USA). Briefly, inoculation of 1:100 of overnight culture, two hours incubation at 37°C and 220 rpm in the orbital shaker followed by the induction with 0.1% L-Rhamnose (10% stock solution in water, AppliChem, Gatersleben, Germany) and four hours incubation at RT and 220 rpm. Subsequently both cultures of each clone were harvested by centrifugation and resuspended in GST-Lysis buffer from the Pierce® GST Spin Purification Kit (Thermo Scientific, Rockford, USA). Then they went through three freeze-thaw cycles with liquid nitrogen and a 15 min sonification step, followed by a 15 min centrifugation at 5000 rpm on 4°C. The supernatant was used for a SDS-PAGE (Invitrogen, Carlsbad, USA). A coomassie stain of the gel showed us which colonies yield best results. We then used these clones according to the protocol described above to prepare 200 ml cultures. The supernatant of these cultures were then used for protein purification with the Pierce® GST Spin Purification Kit (Thermo Scientific, Rockford, USA), following the manufacturer's protocol.

IDH1 enzyme kinetics

The measurement of the enzyme kinetics was performed with an Omega FluoStar (BMG Labtech, Ortenberg, Germany) equipped with a pump system, which was used to start the reaction, by adding 4 µg protein to the reaction mixture. All measurements were performed in

96-well plates (BD Falcon, Franklin Lakes, USA) in a total volume of 100 μ l at 37°C. The reaction mixture consisted of Tris-HCl pH 7.4 (50 mM), MgCl₂ (2 mM), NaCl (10 mM), BSA (0.05%), NADP⁺/NAPDH (10 mM or concentration row), and isocitrate/ α -KG (10 mM or concentration row). As a negative control we used the same mixture, but without isocitrate or α -KG, depending on the K_M to be obtained. After the start of the reaction, a one second double orbital shaking (500 rpm) was used to mix the reaction. Thereafter, every two minutes data was obtained from each well measuring the NADPH production or consumption (Ex. 340 \pm 10 nm, Em. 440 \pm 10 nm), depending on the reaction analyzed. All measurements were done in triplicate and at least 15 data points of each reaction were used for K_M calculation. The mean of three independent measurements and the standard deviation of these is plotted for each K_M.

D-2-hydroxyglutarate

The 2-HG detection was performed with an enzymatic assay developed in our lab. Probe preparations and measurements were performed as described in [16].

D-2-hydroxyglutarate was obtained from Sigma Aldrich (St. Luis, USA) as disodium salt (catalogue number H8378).

Octyl-D-2-hydroxyglutarate was synthesized following the method reported by Xu et al. [12] with slight modifications (detailed protocol for D-2-hydroxyglutarate synthesis, see Additional file 1 and Additional file 2: Figure S1).

Cell lines

All cell lines were achieved from the ATCC and cultured under standard culture conditions (37°C, 5% CO₂) in DMEM medium with 1% Penicillin and Streptomycin and 10% fetal calf serum (all obtained from Gibco® Invitrogen, Carlsbad, USA).

For the generation of IDH1R132H overexpressing cells, IDH1R132H in the destination vector pDEST26 (N-terminal 6x His Tag) was used. LN229 cells were transfected with IDH1R132H in pDEST26 vector by Eugene 6 (Promega, Madison, USA) followed by picking of single cell clones. Single cell clones were selected with 2 mg/ml Gentamycin (Invitrogen, Carlsbad, USA). Clones that survived selection were analyzed for their expression of IDH1R132H by western blot. Clones H3 and H114 were chosen for further analysis, due to their different and stable expression levels of IDH1R132H.

For all experiments with the inducible expression system we used normal cell culture medium, but exchanged normal FCS with 10% Tet system approved FBS (Clontech, Mountain View, USA). To generate an inducible cell line we used the pT-REx-DEST system (Invitrogen, Carlsbad, USA). As a first step we transfected the cell line LN319 with pcDNA6/TR. From this

transfection we generated single cell clones and tested their reliability by introducing EGFP in pT-REx-DEST30. We chose the clone with no GFP expression in tetracycline free media and with the highest expression after induction with 1 μ M Doxycycline (Sigma-Aldrich, St. Luis, USA). This clone 09 (K09) was then used for all further experiments.

For the experimental setup we used LN319 K09 transfected with IDH1 wild type (wt), the different IDH1 mutants, and GFP in pT-REx-DEST30. All cell lines were seeded as described under proliferation analysis. For each cell line two triplicates were seeded. One was induced with 1 μ g/ml Doxycycline, the other one was treated with the comparable amount of solvent (DMSO, Sigma-Aldrich, St. Luis, USA).

To generate cell lines which express IDH1 wt and mutant proteins, we used cDNAs in pMXs-GW-IRES-BsdR and transfected them into HEK293T cells with FuGene®. Cells were subsequently put under selection pressure, by adding 4 μ g/ml Blasticidine S (Sigma-Aldrich, St. Luis, USA). As control we used GFP in pMXs-GW-IRES-BsdR.

Cell line analysis

For cell number analysis CellTiterGlo (Promega, Madison, USA) was used in 96-well plate format. All cells were seeded at a density of 5,000 cells/well and subsequently treated. Measurements were performed at the indicated time points following the manufacturer's protocol.

All migration assays were performed with Ibidi Culture-Inserts. In each well of the insert, 50,000 cells were seeded. After 24 h, the cells were treated with 10 μ g/ml Mitomycin C (Sigma-Aldrich, St. Luis, USA) for 2 h, to avoid the influence of proliferation in the assay. Afterward, the inserts were removed leaving a gap of 500 μ m \pm 50 μ m. Cells were washed with PBS once and thereafter normal medium was applied. This medium was subsequently substituted with 2-HG at the given concentrations. The gap was microscopically documented at the start and the indicated time points. The pictures were analyzed with TScratch (<http://chaton.ethz.ch/software/>) and the area closed after the indicated time point was plotted in the graph.

The soft agar assays were performed in 6-well plates. The wells were prepared with a 1% agar solution (Agar noble, US Biological, Salem, USA) as base agar. On top of this, 5,000 cells in 0.35% agar were seeded and grown for 14 days. The cells were stained with 0.00025% crystal violet solution and colonies > 1 mm² were counted.

Results

Distribution of IDH1 mutations in gliomas

Including data from a previous study [2] and on ongoing routine analysis of all diffuse astrocytoma and oligodendroglioma in the Department of Neuropathology at the University Heidelberg, the present series of IDH1

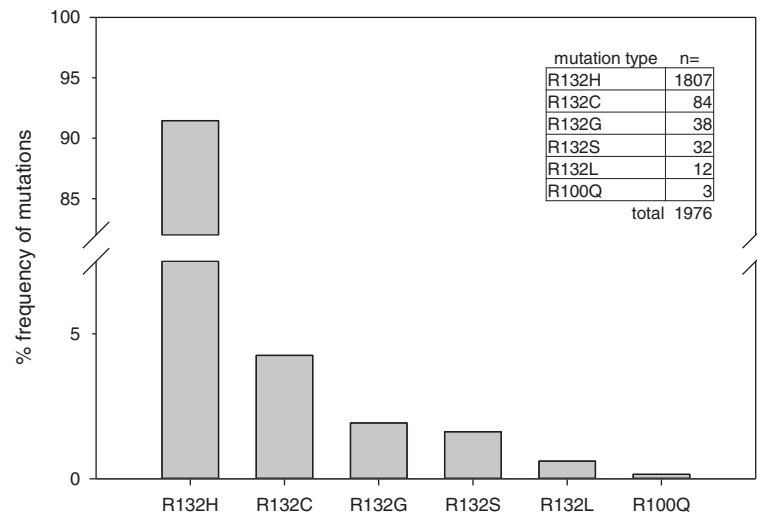


Figure 1 Distribution of IDH1 mutation types. Distribution of mutation types in the Heidelberg series of 1,976 diffuse astrocytomas and oligodendrogliomas with *IDH1* mutations.

mutated gliomas comprises tumors from 1,976 patients. These 1,976 tumors represent only the immunohistochemical and sequence verified *IDH1* mutant cases out of the whole collection of glial tumors in our department. The frequency of *IDH1* mutations in these tumors decreases in the order R132H (91.5%), R132C (4.3%), R132G (1.9%), R132S (1.6%), R132L (0.6%) and R100Q (0.3%). Incidences of the mutations are given in Figure 1.

2-HG in tumor tissue

We determined the amount of 2-HG in glioma tissue. The limiting step of our analyses was the low incidence

of *IDH1* mutations in gliomas other than R132H. Sufficient formalin fixed and paraffin embedded tissue was available from 9 tumors with R132C, 6 tumors with R132G and 1 each with R132S and R132L. Due to loss of 2-HG during the processing up to paraffin embedding [13], only approximate values for 2-HG could be determined. Lowest 2-HG levels were detected in tumors with R132H followed by R132C and then R132G. The single tumors with R132S and R132L mutation also exhibited high levels of 2-HG. No 2-HG was detected in 5 control glioma samples without *IDH1* mutation. The data is shown in Figure 2.

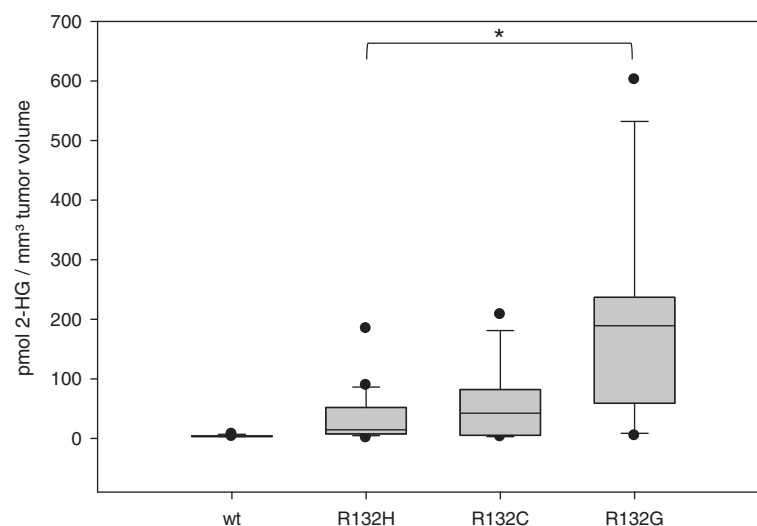


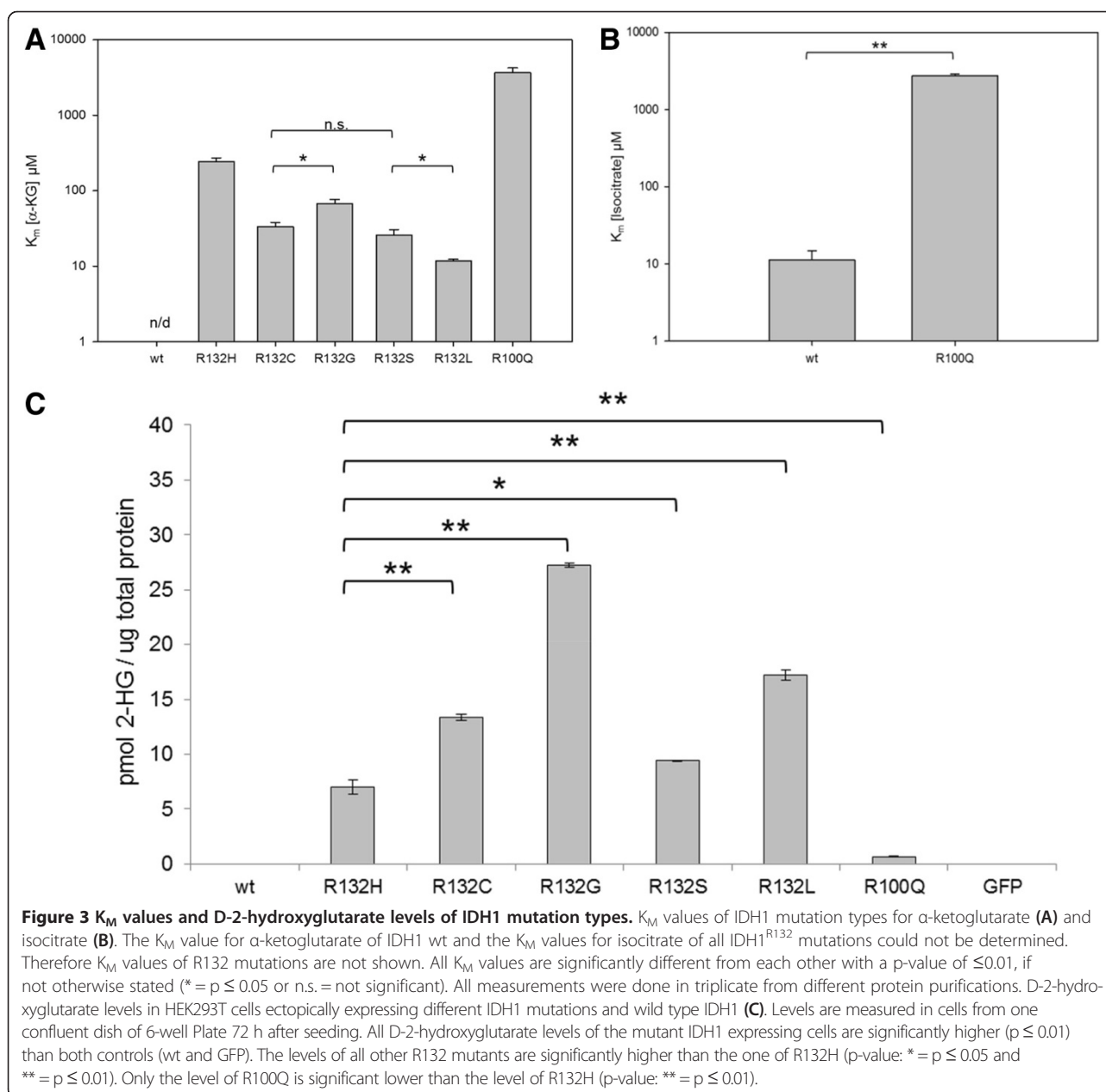
Figure 2 D-2-hydroxyglutarate levels in FFPE tissue. D-2-hydroxyglutarate levels in FFPE tissue from diffuse astrocytomas and oligodendrogliomas with *IDH1* mutations. Shown are the levels of 6 *IDH1* wt tumors, 25 with *IDH1*R132H, 8 with *IDH1*R132C and 7 with *IDH1*R132G. All mutant tumors show significant (** = $p \leq 0.01$) higher 2-HG levels than wildtype tumors, but only the difference between R132H and R132G mutation was statistically significant (* = $p \leq 0.05$).

Enzyme kinetics

Recombinant expression of proteins with IDH1 mutations followed by purification enabled the determination of Michaelis constants for each mutant protein with the substrate α -KG. A high K_M of α -KG was observed for the R132H mutation (243.7 μ M) whereas lower K_M 's were seen for R132G (67.4 μ M), R132C (33.7 μ M), R132S (26.1 μ M) and R132L (11.8 μ M). Data is shown in Figure 3A.

The very rare R100Q mutation exhibited a high K_M of (3671.1 μ M). This was the only mutated IDH1 protein for which a Michaelis constant for isocitrate was

detected ($K_M = 2724.0 \mu$ M) (Figures 3A and 3B). To further validate this data, we transfected HEK293T cells with all different IDH1 variants and determined the cellular 2-HG level (Figure 3C). As expected the R132C mutant produced more 2-HG than R132H mutant. Also, the other mutants R132G, R132S and R132L all produced more than R132H. However, the 2-HG concentration measured did not exactly correspond to the K_M values determined. The 2-HG concentration from R100Q was as low as expected, ranging below that from R132H. To exclude the possibility that the different 2-HG concentrations are due to differential expression



of the respective mutant proteins or different protein stabilities, we performed a western blot demonstrating comparable protein quantity (Additional file 3: Figure S5).

Cell viability dependence on 2-HG concentration

Viability upon exogenous exposure to 2-HG was tested using immortalized human astrocytes, HEK293T cells, and seven glioma lines (LN18, LN229, LN319, A172, U87, U373, T98G) by addition of cell permeable octyl-2-HG to medium [12]. To determine the concentration range to be used, we calculated the molar 2-HG concentrations in the tumor specimens from previous reports [17,18] and our tumor material. For converting concentrations given in $\mu\text{mol/g}$ to molarities we assumed brain tumor tissue density to be similar to normal human brain tissue measuring $1.054 \pm 0.014 \text{ g/cm}^3$ [19]. On this basis, 2-HG concentrations in brain tumor tissues range from 0.1–126 mM. We, therefore, added cell permeable octyl-2-HG to cell medium in concentrations

ranging from 100 μM to 150 mM. All cell lines challenged perished at 2-HG concentrations of 7.5 mM and above. Therefore, only data using octyl-2-HG concentrations ranging from 250 μM to 7.5 mM are shown in Figure 4A.

Due to the high toxicity of octyl-2-HG we switched to the less cell permeable sodium salt of 2-HG, because we aimed for more physiological concentrations in our experiments. We added this 2-HG to HEK293T, human astrocytes and LN229 in concentrations ranging from 500 μM to 50 mM. Concentrations at the low end from 500 μM to 1 mM had no significant effect on proliferation. From 2.5 mM to 50 mM, increasing toxicity was observed with LN229 being more resistant than HEK293T and human astrocytes (Figure 4B).

In order to test the effect of endogenous 2-HG we transfected LN229 with pDEST26 containing an IDH1R132H construct. Two clones H3 and H114 were established by single cell cloning, both stably producing 2-HG, albeit

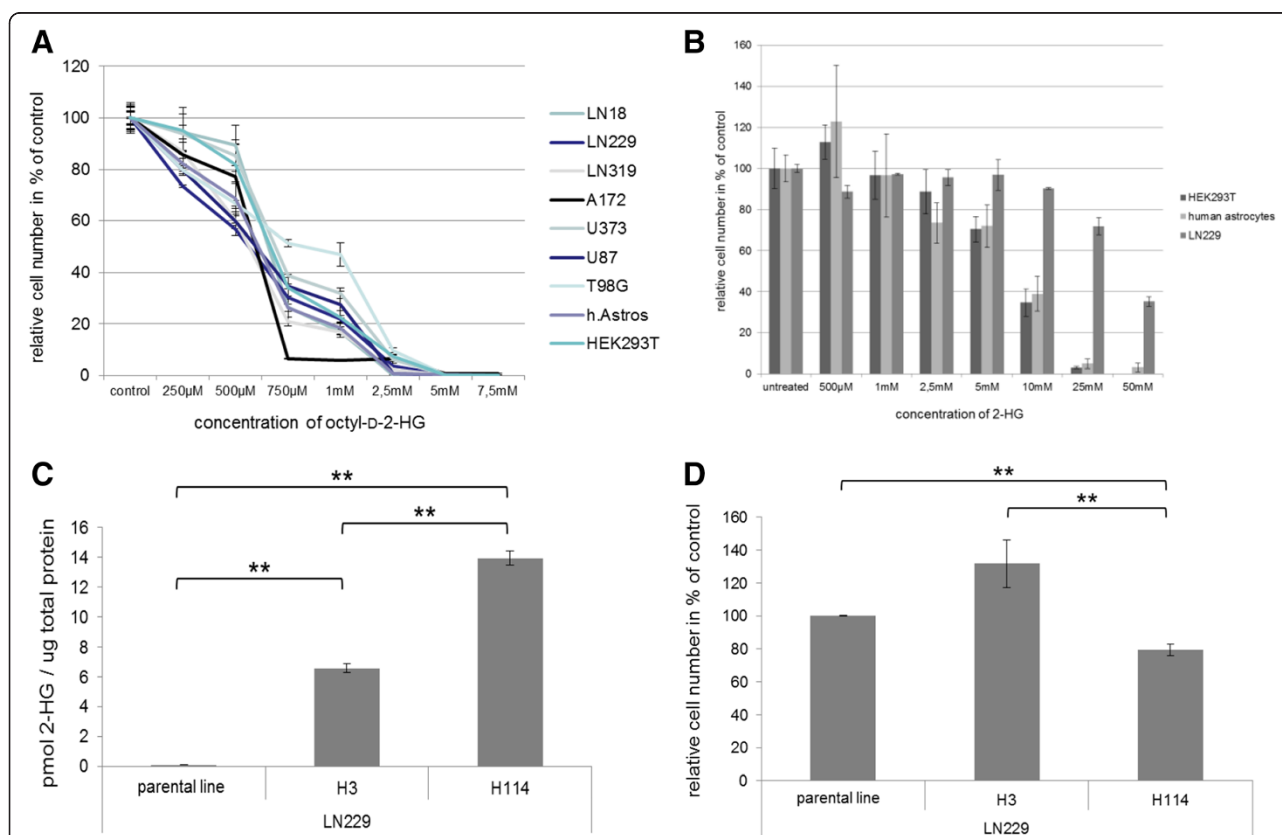


Figure 4 D-2-hydroxyglutarate toxicity and analysis of two LN229 sublines. Octyl-D-2-hydroxyglutarate mediated toxicity in different cell lines (A). D-2-hydroxyglutarate mediated toxicity in different cell lines (B). Toxicity was measured 48 h after treatment. Plotted are the relative cell numbers in % compared to the corresponding untreated control. D-2-hydroxyglutarate levels in LN229 glioma cell line and two LN229 glioma cell subclones H3 and H114 overexpressing different protein levels of R132H (C). All D-2-hydroxyglutarate levels are significantly different from each other ($p \leq 0.01$). Proliferation of LN229 glioma cell line and of two LN229 glioma cell subclones H3 and H114 overexpressing different protein levels of R132H (D). The relative cell numbers were determined 48 h after seeding and are plotted in % compared to the corresponding control. The enhanced proliferation of subclone H3 is not significant compared to the parental line, but the reduced proliferation of subclone H114 is statistically significant compared to both other lines ($p \leq 0.01$).

at different levels. Clone LN229 H3 contained 6.58 pmol 2-HG/ μ g total protein, a concentration not significantly affecting proliferation. In fact, a trend for higher proliferation was noted. In contrast, clone LN229 H114 containing 13.94 pmol 2-HG/ μ g total protein exhibited a significant reduction of proliferation (Figures 4D).

Concentration dependent toxicity of different IDH1 mutations

We transfected in LN319 and HEK293T cells with pcDNA6/TR and selected single colonies. This was done to have cell lines with high expression of TR (tet repressor) sufficient for silencing pT-Rex-DEST30 prior to induction with Doxycycline.

We cloned IDH1 wt, all the R132 mutants and the R100Q mutant in pT-Rex-DEST30, and transfected these constructs into LN319 and HEK293T containing pcDNA6/TR. Then, the expression of the respective mutant IDH protein was induced.

Induction of R132H had no effect on proliferation in LN319, but in HEK293T it lead to a reduction in growth. Induction of all the mutations with lower K_M for α -KG resulted in reduced cell growth in both lines. As expected, expression of the R100Q mutation, producing low levels of 2-HG, also had no effect on growth in LN319. Neither expression of wt IDH1 protein nor of GFP affected cell growth in LN319. Results of growth kinetics for the different transfectants are shown in Figure 5.

Effect of 2-HG concentration on migration and colony formation

Wound healing assays were performed employing LN229 and different concentrations of 2-HG. In addition, we

performed this assay employing LN229 H3 and LN229 H114 cells with ectopic expression of mutant R132H IDH protein at different levels. Addition of 2-HG at low concentration of 100 μ M resulted in enhanced migration, which was less pronounced at higher concentrations of 10 mM. LN229 H3 showed a migration rate higher than the parental LN229 while LN229 H114 did not differ from LN229 in this respect (Figure 6A).

Colony formation in soft agar was augmented upon exposure to 2-HG in moderate concentrations. Concentrations of 100 μ M and 10 mM 2-HG in medium both resulted in an increase of colonies by 61% and 59%, respectively. LN229 H3 containing 6.58 pmol 2-HG/ μ g total protein exhibited a 3-fold increase in colony formation. In contrast, LN229 H114 containing 13.94 pmol 2-HG/ μ g total protein did not form significantly more colonies than LN229 without treatment (Figure 6B).

Discussion

The IDH1 mutation of the R132H type by far outnumbers the other IDH1 mutations in diffuse astrocytic and oligodendroglial tumors. In order to provide a hypothesis for this lopsided distribution, we characterized biochemical features of different IDH1 mutations, analyzed native tumors and performed experiments addressing the effects of different mutations on *in vitro* systems.

Common to all IDH1 mutations is acquisition of a neo-enzymatic activity with the ability to convert α -KG to 2-HG. Thus, we determined the Michaelis constants of mutated IDH proteins for the substrate α -KG. We detected the highest K_M for α -KG in protein carrying the R132H mutation followed by R132G, R132C, R132S and R132L (Figure 3). While increased 2-HG levels are

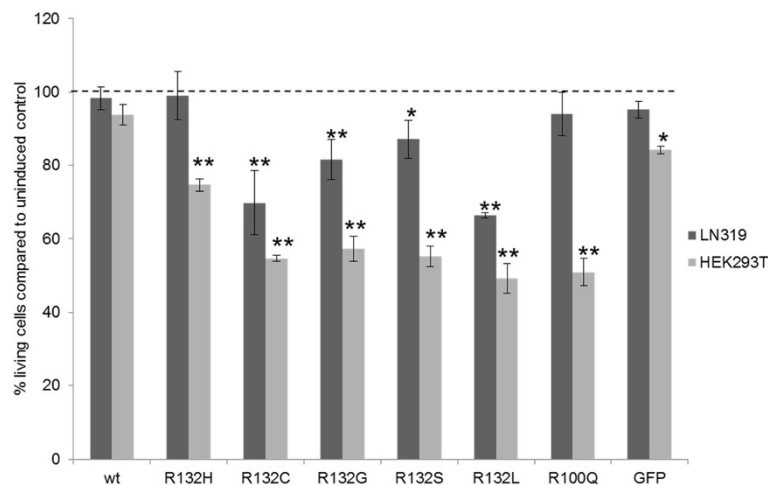


Figure 5 Proliferation analysis of LN319 glioma and HEK293T cells with and without induction of mutant IDH expression. Cell viability of LN319 glioma and HEK293T cells bearing tetracycline inducible constructs of different IDH mutations and controls 72 h after induction. Shown are the relative cell numbers compared to the corresponding uninduced controls. The significant differences are depicted in the figure with their corresponding to their p-values (* = $p \leq 0.05$ and ** = $p \leq 0.01$).

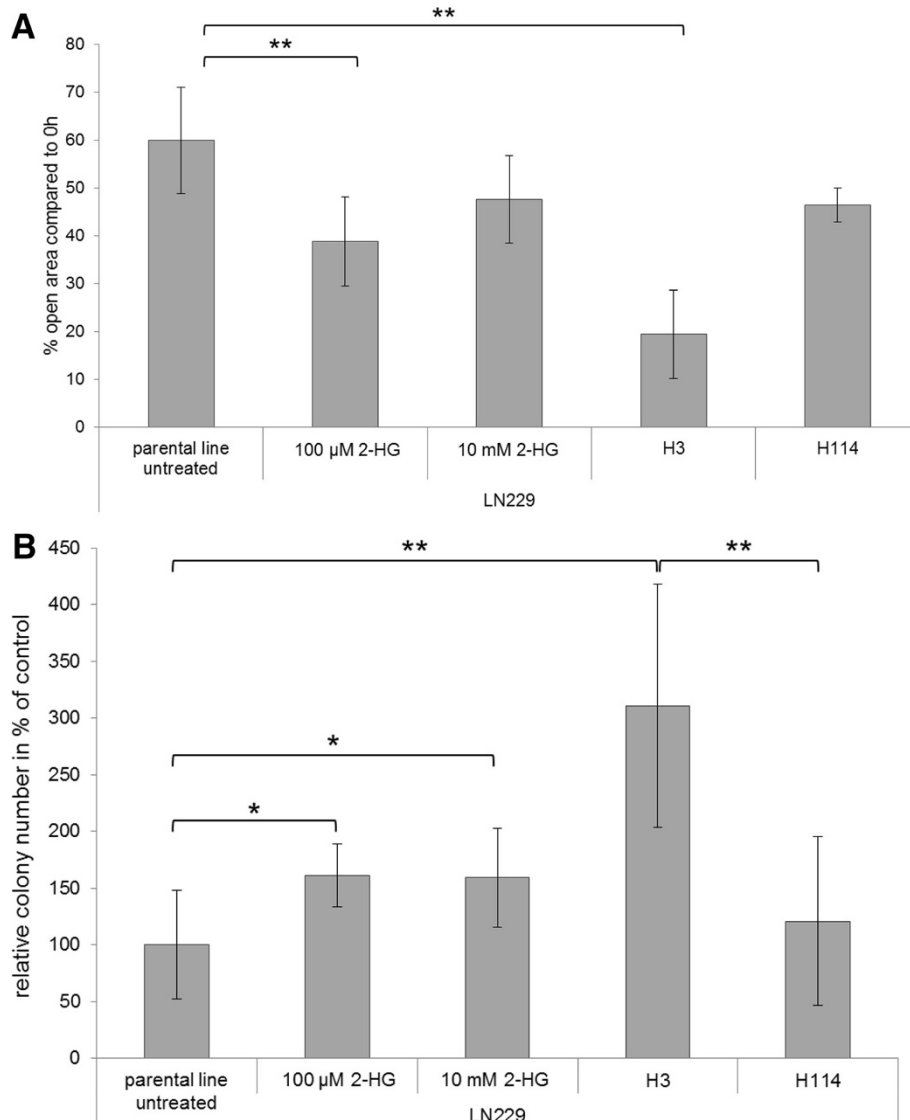


Figure 6 Migration and soft agar analysis of LN229 treated with D-2-hydroxyglutarate and LN229 subclones. Migration of LN229 and subclones H3 and H114 after 72 h. LN229 cells were treated with 100 μM or 10 mM 2-HG. Subclones H3 and H114 expressed R132H protein in different amounts (A). Plotted is the open area after 24 h of incubation. Treatment with 100 μM 2-HG or ectopic expression of low amounts of R132H protein (subclone H3) increased migration speed significantly (** = $p \leq 0.01$). Colony formation in soft agar of the same cells in the same setting (B). Shown are the relative colony numbers compared to the untreated parental line (100%). The 2-HG treatment could significantly (* = $p \leq 0.05$) increase colony formation independent of the used concentration. An even stronger increase (** = $p \leq 0.01$) could be measured for subclone H3, but no difference could be detected for subclone H114.

expected to provide a selection advantage for early tumor cells, it should be kept in mind that high 2-HG levels may be toxic. Thus, positive discrimination for a mutation type resulting in a protein with moderate activity for features beneficial at low but potentially deleterious at high dose is well compatible with a selection process.

A beneficial effect of moderately increased 2-HG on proliferation (cell line LN229 H3, 6.58 pmol 2-HG/μg total protein) is demonstrated in Figure 4D. In contrast, strongly increased 2-HG levels (cell line LN229 H114,

13.94 pmol 2-HG/μg total protein) did not favor proliferation. Similarly, migration could be increased for LN229 by 100 μM 2-HG in the medium and slightly less by 10 mM 2-HG in the medium. Moreover, the cell line LN229 H3 with moderately induced 2-HG production exhibited increased migration compared to LN229. LN229 H114, with higher 2-HG production, did not migrate significantly faster than LN229 (Figure 6A). Likewise, colony formation was strongly supported by moderate 2-HG concentrations in the LN229 and subclone LN229 H3. In contrast, LN229

H114, producing high levels of 2-HG, exhibited no benefit (Figure 6B).

To test the effect of 2-HG on viability of cells we added 2-HG, or the more cell permeable octyl-2-HG [12], to culture medium. In our models employing human astrocytes and HEK293T, LD₅₀ for 2-HG was determined for concentrations between 5 mM and 10 mM while the LD₅₀ for LN229 was approximately 50 mM (Figure 4B). In the same cell lines and several more, the LD₅₀ for octyl-2-HG was reached at approximately 750 μM (Figure 4A). Moreover, transfection and induction of the different IDH1 mutations into LN319 cells demonstrated no influence on proliferation for R132H; however, the other mutations with lower K_M for α-KG inhibited proliferation. These findings demonstrated considerable cell toxicity of 2-HG.

To further support this hypothesis we analyzed the amount of 2-HG in formalin fixed and paraffin embedded brain tumors harboring different IDH mutations. The limiting step of our analyses was the low incidence of IDH1 mutations other than R132H in gliomas. The detection of moderately increased 2-HG levels in tumors with the IDH1R132H mutation and higher levels in the rare mutation types matched well with the different affinities for α-KG of the respective mutations.

Thus, in glioma cells we demonstrate beneficial effects of moderate 2-HG concentrations for proliferation, migration and colony formation and toxic effects on these readouts for high 2-HG concentrations. This would well match a model favoring an IDH1 mutation with an intermediate K_M for α-KG.

These findings may explain the strong preponderance of the IDH1R132H mutation type in glioma.

Both exo- and endogenous mechanisms affect types of mutation in DNA. Sporadic primary brain tumors in humans have not been shown to be associated with typical exogenous DNA damage such as ultraviolet-induced DNA-damage or that following exposure to carcinogens. It is unresolved whether accumulation of distinct mutations such as IDH1 originates from an endogenously mediated increase or a specific failure of repairing this distinct alteration. In low grade glioma, cytosine to thymidine transitions constitute the most frequent mutation type on the single nucleotide level [20]. R132H facilitated by a CGT to CAT change corresponds to a C to T transition on the reverse strand. On the other hand, R132C facilitated by CGT to TGT is based on this transition on the coding strand. Neglecting potential repair of the C to T transition on the reverse strand during replication, R132H and R132C could be expected to occur with comparable frequency. This holds true for these mutations in acute myeloid leukemia [5,21]. In contrast, IDH1R132H dominates in diffuse gliomas and R132C appears to be the most frequent mutation in chondrosarcoma [6,7] and intrahepatic cholangiocarcinoma [8,22]. Thus, the strong bias for

IDH1R132H mutation in astrocytoma and oligodendroglioma may support a selection bias for this alteration.

Conclusion

We determined significantly different enzymatic activities for distinct IDH1 mutations and provide a selection based hypothesis for the preponderance of the IDH1R132H mutation in astrocytoma and oligodendroglioma.

Additional files

Additional file 1: d-2-hydroxyglutarate synthesis.

Additional file 2: Figure S1. Synthetic route to octyl-D-2-HG.

Additional file 3: Figure S5. Western blot analysis of overexpression cell lines.

Competing interests

AvD, JB and SP hold a patent for the enzymatic D-2-hydroxyglutarate detection assay. All other authors declare that they have no competing interests.

Authors' contributions

Conceived and designed the experiments SP. Generated mutant proteins and vectors SP, inducible cell line JL SP, over expression cell lines SW AB SP. Performed soft agar assay and migration assay LS, proliferation analysis SP, enzyme kinetics AB JL JB, 2-HG detection assay AB JB, octyl-2-HG synthesis AM, database analysis AvD, statistical analysis LS SP. Wrote the manuscript AvD LS SP. All authors read and approved the final manuscript.

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